**AN OVERVIEW OF AZOLE ANTIFUNGALS**

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**Abstract:**

As the number of individuals with immune system suppression has grown in recent years, the danger of human fungal infections has significantly increased. Fortunately, there has been a rapid advancement in the treatment of fungal infections. It was discovered that combining already available azoles with additional antifungals is probably going to increase their efficacy. But after receiving antifungal medication, a number of circumstances might cause therapeutic failure or recurrence. These variables relate to the various properties of the antifungal(s) employed. Therefore, professionals should thoroughly research the various properties of antifungals in order to prevent these issues and further to make the best use of the antifungals.

The goal of the current study was to identify variations in the pharmacology, pharmacokinetics, spectrum of activity, toxicity, and possible medication interactions of the related azoles.

The azoles of interest are sufficiently diverse in terms of their pharmacology, pharmacokinetics, spectrum of activity, safety, toxicity, and potential drug interactions, according to the current review, which enables medical professionals to distinguish between these agents based on their individual properties when designing a patient's therapy.

**Keywords:** Human fungal infections, Azoles of interest, Different characteristics and development in the antifungal chemotherapy.

**Introduction:**

Infectious disease is a condition in which the infecting agents do cause the body to react, which results in clinically manifest signs and symptoms. Humans and the surrounding microorganisms coexist peacefully, but an infection may arise from the microorganisms when the defence system is compromised or the concentration of pathogens reach an exceptionally high density. Infectious illnesses have been brought on by bacteria, viruses, parasites, fungus, prions, worms, and parasitic organism among others. Infections brought on by bacteria were formerly the most dreaded, but as methods of treating bacterial infections in patients became more effective, fungus have now replaced bacteria as the most dangerous pathogens [1]. Molds and yeasts are the two most common types of fungi. Mold colonies are made up of filamentous threads called hyphae, whereas yeasts are normally made up of a single, tiny, oval cell. Some fungi are dimorphic; depending on the external environment, such as temperature, they may either exist as yeasts or molds. Molds and yeasts are the two most common types of fungi. Mold colonies are made up of filamentous threads called hyphae, whereas yeasts are normally made up of a single, tiny, oval cell. Some fungi are dimorphic; depending on the external environment, such as temperature, they may either exist as yeasts or molds. [2, 3] Even while fungi are common in the environment, only a few species are frequently discovered in close proximity to people who might spread illness. The majority of fungi that primarily affect immunosuppressed people are frequently categorised as opportunistic pathogens (Candida and Cryptococcus), while a small number of fungi that are responsible for causing disease in healthy people are considered to be true pathogens (Histoplasma and Paracoccidioides) [4]. But it's clear that some opportunistic fungal infections can also lead to illness in otherwise healthy people (Candida vaginitis or Cryptococcus gattii outbreaks) [5,6] High morbidity and mortality are characteristics of invasive fungal infections, which are still challenging to identify, prevent, and treat despite becoming increasingly prevalent. [7]

**Types of fungal infections**

1. **Topical/superficial disease caused by fungal pathogens:**

The outermost layers of the skin, nails, hair, and mucous membranes can develop superficial fungal infections. [8]

**Dermatophytosis**

Organisms that consume keratin are called dermatophyte fungus [9].

The stratum corneum of the epidermis and tissues made of keratin, such as the hair and nail, are infected by dermatophytes. Most superficial fungal infections are caused by Trichophyton spp., Microsporum spp., and Epidermophyton spp., while some yeast and other non-dermatophyte moulds can also be the culprits. [10]

**Tinea pedis**

The primary pathogens responsible for tinea pedis are Trichophyton rubrum, Trichophyton mentagrophytes, and Epidermophyton floccosum. Tinea pedis is a dermatophyte infection of the foot that mostly affects the toes and sole. This infection, which is the most prevalent dermatophyte fungal illness to afflict men [11], affects between 5 and 30 percent of the population. People who have tinea pedis may be more prone to subsequent bacterial infections, such as those caused by Group a streptococcus. [12]

**Tinea corporis**

T. rubrum and T. mentagrophytes, which affect the neck, trunk, and extremities, are two of the main causal organisms for tinea corporis. A finely defined circular lesion with erythema, scaling, and tiny blisters or pustules at the border is the characteristic of a tinea corporis lesion. The lesion typically has a diameter of 5 cm. Domesticated animals including cats, dogs, hamsters, and guinea pigs frequently transfer the fungus to people [11].

**Tinea capitis**

Trichophyton tonsurans is the most common cause of this illness, which often affects children and manifests as baldness and scaling on the scalp [13, 14].

**Tinea unguium or onychomycosis (nails)**

The main causes of onychomycosis are T. rubrum and T. mentagrophytes dermatophytes, which account for over 90% of toenail infections and 50% of fingernail infections [15–16].

1. **Malassezia infection**

Malassezia spp. are widespread surface commensals of oily skin, including the scalp and chest, and they are linked to folliculitis, seborrheic dermatitis, and pityriasis versicolor [17].

1. **Malassezia folliculitis**

A rash on the upper back and shoulders caused by Malassezia folliculitis can itch and look like acne [18].

1. **Subcutaneous Infection**

Although they can spread, subcutaneous mycoses typically only affect the dermis and subcutaneous tissues.

**Sporotrichosis**

The most common subcutaneous infection is sporotrichosis, which is brought on by the dimorphic fungus Sporothrix schenckii . The fungus is present in soil and plants, and it typically injures farmers and gardeners, particularly those who take care of roses. It is a small cutaneous or subcutaneous lesion that has the potential to grow and spread through the lymphatic system. The condition known as lymphocutaneous sporotrichosis is not life-threatening [19].

**Chromoblastomycosis**

The Dematiaceae family of fungi, which includes Fonsecaea pedrosoi, Cladosporium carrionii, Fonsecaea compacta, Phialophora verrucosa, and Rhinocladiella aquaspersa, are responsible for the chronic cutaneous or subcutaneous fungal infection known as chromoblastomycosis [26]. They can be found in wood, vegetable waste, and soil. Raised, crusty skin lesions are the symptoms.

**Chronic mucocutaneous candidiasis**

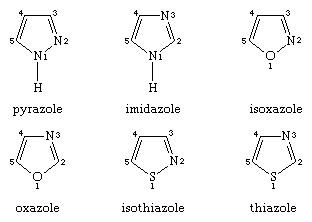
A uncommon illness called chronic mucocutaneous candidiasis involves persistent C. albicans infection of the mucous membranes, which may spread to the skin and nails. Though the underlying problem is still not well understood, the disorder is linked to defective cell-mediated responses to Candida [27,28]. White fissured lesions, hyperkeratotic, granulomatous, and vegetative lesions, as well as an autosomal recessive characteristic linked to endocrine problems such hyperparathyroidism are some of the signs.

**Systemic fungal infections**

The endemic or dimorphic mycoses are two separate categories of systemic fungi diseases. Unlike opportunistic mould and yeast infections, which are saprophytes and only invade an immunocompromised host, these infections are brought on by real pathogenic fungi [36,37]. These infections pose a serious risk to health and are frequently fatal. Patients with solid organ transplants who use immunosuppressive drugs to reduce the risk of rejection are more vulnerable to systemic fungal infections [2,3].

**Classification of Azole group**

According to fig. 1, the azoles are subclasses of five-membered nitrogen heterocyclic ring compounds that contain at least one additional nitrogen, sulphur, or oxygen atom. The parent compounds have two double bonds and just one ion pair of electrons from each heteroatom in the ring, and they are aromatic. In azoles, the heteroatom that is not a component of a double bond is numbered first, immediately by the other heteroatom [20].



**Fig. 1: The different basic structural rings of azoles**

Azoles are classes of fungi static substances having wide-ranging action. Imidazole’s and triazoles are the two groups into which they are divided. Each group's members are structurally related, and changes to side-chain structure determine both the antifungal activity and level of toxicity.[21]

**Pharmacology of azole:**

**Mechanism of action:**

Fluconazole, itraconazole, ketoconazole, posaconazole, and voriconazole are some of the azoles with systemic action. By inhibiting CYP-dependent 14-demethylase, which is required for converting lanosterol to ergosterol, the azoles have a fungistatic action. The stability of the fungal cell membrane depends on ergosterol, and inhibiting its production compromises membrane integrity [22].

Additionally, the triazoles indirectly target several stages of the process for ergosterol production. For instance, in C. albicans that is fluconazole-sensitive, fluconazole only partially inhibits the synthesis of ergosterol and totally blocks the synthesis of obtusifoliol, but voriconazole completely inhibits the production of both ergosterol and obtusifoliol 5. The enzyme 3- ketoreductase, which C. neoformans uses to convert the 3-ketosteroid obtusifolione to obtusifoliol, may likewise be inhibited by itraconazole and fluconazole. [23]

**Spectrum of activity**

Azoles have a wide range of action against moulds and yeasts. However, as this therapeutic class grows, variations in the distinct medicines' activity spectra become apparent. The discrepancy in the spectrum of action may be explained by the different ways that different species inhibit 14-demethylase and secondary targets. Although they are infrequently employed for this purpose, azoles also have some antibacterial properties. The majority of important fungi and yeasts are resistant to miconazole's broad antifungal range.[24] Sensitive organisms include Blastomyces dermatitidis, Coccidioides immitis, Paracoccidioides, Histoplasma capsulatum, Candida species, including (C. Krusei, C. Inconspicua, C.Albicans, C. Lusitaniae, C. Glabrata, C. Guilliermondii, C.Tropicalis, C. and C. Parapsilosis), Cryptococcus neoformans, and Aspergillus fumigatus [24] Similar to miconazole in terms of its antifungal range, ketoconazole is more potent against C immitis and a few other yeasts and fungi. [25] C. neoformans and Coccidioides immitis are both susceptible to fluconazole, while Aspergillus species are not species of Fusarium and the zygomycosis agents. [25] Itraconazole is typically fungistatic to numerous yeasts and exhibits fungicidal action against filamentous fungi and some strains of C. neoformans. When used against the majority of fluconazole-susceptible and fluconazole-resistant Candida species, it has moderate to high activity (except C. Glabrata). [25]

**Pharmacokinetics of Azole**

**Absorption, distribution, biotransformation and excretion [29, 30]**

The azoles are weak lipophilic bases chemically. After oral administration, they have an excellent relative or absolute bioavailability (except the capsule form of itraconazole). A reduction in stomach acidity might decrease the bioavailability following delivery via oral route of the azoles, with the exception of fluconazole, which requires an acidic environment to dissolve. There are contradicting accounts, however it seems that taking the medication with food speeds up absorption. Elevated gastric pH has a considerable impact on how well the solid oral dose forms of itraconazole and ketoconazole dissolve in the stomach [31].

Fluconazole requires substantially less biotransformation to be removed from the body than the other azoles since it is hydrophilic and highly soluble in water. Itraconazole and voriconazole have a low solubility in aqueous solution and are very lipophilic [32]. The main oxidative drug-metabolizing enzyme in humans, CYP3A4 is inhibited by azoles [33]. The affinities of the azoles for this enzyme vary. Additionally, fluconazole and voriconazole both inhibit CYP2C9/19, and fluconazole also blocks the UGT2B7 pathway [34]. The main method of removal is by hepatic metabolism [32]

To be excreted from the body, azoles need a lot of oxidative (CYP) metabolism [32]. Only 2-4% of a dosage given orally shows up in urine unaltered. Itraconazole undergoes active metabolite metabolism, which may greatly increase the drug's antibacterial effectiveness. 20% of the metabolites are removed in the urine, and the biliary route accounts for the majority of excretory pathways (>80%) [34].

Several transport proteins, which are expressed in tissues all across the human body, help with drug disposition. The interactions of the azoles with transport proteins differ [35, 36]. With measurable amounts in saliva, milk, and cerumen, the azoles seem to be broadly dispersed throughout the body. With the exception of fluconazole, which reaches 50–90% of plasma concentrations, cerebral spinal fluid penetration is relatively low.

With the exception of fluconazole, the majority of azoles are strongly protein-bound in the bloodstream (>95%), mostly to albumin [35, 36]. The liver, adrenal glands, lungs, and kidneys have the largest quantities of azoles. The most well-known efflux transport protein, P-glycoprotein, interacts with itraconazole and ketoconazole [35, 37].

Itraconazole and ketoconazole interact with the breast cancer resistance protein (BCRP), a different transporter [35, 36]. Although the relevance of these interactions with BCRP has not yet been fully clarified, they may help to partially explain several interactions that cytochrome P450 interactions alone were previously unable to effectively explain [35, 36].

The half-life of the azoles' rate of elimination appears to be dose-dependent: the higher the dosage, the longer the half-life [35, 36]. A biphasic elimination pattern is also seen, with fast clearance in the first 1-2 hours and a gradual drop during the next 6–9 hours. Time to effectiveness may take longer than with medications like amphotericin B because of the extended half-life and mode of action (impaired production of the fungal cell membrane).

**Adverse effect and toxicity**

The azoles administered orally have a number of adverse effects. Cardiopulmonary (hypotension, peripheral/pulmonary edema), CNS (dizziness, headache, seizure), dermatologic/hypersensitivity (anaphylaxis, eosinophilia, pruritus, rash), electrolyte disturbances (hypokalemia), gastrointestinal (abdominal pain/dyspepsia, diarrhea, disguise, nausea/vomiting), hematological (anemia, myelosuppression (alopecia, fever). But it's possible to develop nausea, vomiting, and hepatic impairment, especially with ketoconazole [38].

Particularly with ketoconazole, altered testosterone and cortisol metabolism has been documented [38]. Ketoconazole medication may cause reproductive issues in dogs. Vision disturbances are one of the side effects of voriconazole that can occur in people [39]. Congestive cardiac failure has been linked to the use of itraconazole [40].

**Intraction**

Since many azoles are lipophilic, interactions involving their biotransformation and disposal are involved. Several diverse processes contribute to the azoles' drug interactions. Through a variety of mechanisms, including pharmacodynamic, pH, complexation and electrostatic interactions, CYP and P-glycoprotein, these substances can interact with medications [41]. The azoles' pharmacokinetic interactions are a result of their physicochemical characteristics. Itraconazole and ketoconazole are susceptible to metabolic and pH-based interactions. Agents that are cationic, elevate stomach pH, or are lipophilic CYP3A4 substrates with low oral bioavailability are drugs that are likely to interact with these azoles [41]. All azoles are weak bases, and weakly basic substances dissolve more slowly at higher pH levels. As a result, changes in stomach pH can affect how well azoles like itraconazole capsules are absorbed. [43]. Concurrent use of cimetidine, ranitidine, anticholinergic drugs, or stomach antacids reduces the absorption of all azoles except fluconazole. [43]

**Conclusion**

In recent years, there has been an accelerated growth in the field of fungal infection treatment. The majority of clinically significantSubcutaneous Infection, topical infection, Malassezia folliculitis have responded exceptionally well to treatment using drugs from the azole group. To utilise these drugs effectively, clinicians must be aware of the variations in toxicity and the possibility of drug-drug interactions.

**References:**

1. Wisplinghoff H, Seifert H, Wenzel RP, Edmond MB. Current trends in the epidemiology of nosocomial bloodstream infections in patients with hematological malignancies and solid neoplasms in hospitals in the United States. Clin Infect Dis 2003; 36:1103-10.
2. Groll AH, Shah PM, Mentzel C, Schneider M, Just-Nuebling G, Huebner K. Trends in the postmortem epidemiology of invasive fungal infections at a university hospital. J Infect 1996; 33:23-32.
3. Denning DW, Evans EG, Kibbler CC, Richardson MD, Roberts MM, Rogers TR, et al. Guidelines for the investigation of invasive fungal infections in haematological malignancy and solid organ transplantation. British Society for Medical Mycology. Eur J Clin Microbiol Infect Dis 1997;16:424-36.
4. Brown GD, Denning DW, Gow NA, Levitz SM, Netea MG, White TC. Hidden killers: Human fungal infections. Sci Transl Med 2012;4:165rv13
5. Fidel PL Jr., Barousse M, Espinosa T, Ficarra M, Sturtevant J, Martin DH, et al. An intravaginal live Candida challenge in humans leads to new hypotheses for the immunopathogenesis of vulvovaginal candidiasis. Infect Immun 2004;72:2939-46.
6. Byrnes EJ 3rd, Li W, Lewit Y, Ma H, Voelz K, Ren P, et al. Emergence and pathogenicity of highly virulent Cryptococcus gattii genotypes in the Northwest United States. PLoS Pathog 2010;6:e1000850.
7. Talaviya S, Majmudar F. Recent developments in antifungal agents. Int J Pharm Pharm Sci 2012;4 Suppl 4:4-10.
8. Detandt M, Nolard N. Fungal contamination of the floors of swimming pools, particularly subtropical swimming paradises. Mycoses 1995;38:509-13.
9. Canavan TN, Elewski BE. Identifying signs of tinea pedis: A key to understanding clinical variables. J Drugs Dermatol 2015;14:s42-7.
10. Evans EG. Tinea pedis: Clinical experience and efficacy of short treatment. Dermatology 1997;194 Suppl 1:3-6.
11. Arnold HL, Odom R, William J. Andrews’ Diseases of the Skin. 8th ed. Philadelphia: W.B. Saunders; 1990
12. Semel JD, Goldin H. Association of athlete’s foot with cellulitis of the lower extremities: Diagnostic value of bacterial cultures of ipsilateral interdigital space samples. Clin Infect Dis 1996;23:1162-4
13. Fuller LC, Barton RC, Mohd Mustapa MF, Proudfoot LE, Punjabi SP, Higgins EM. British association of dermatologists’ guidelines for the management of tinea capitis 2014. Br J Dermatol 2014;171:454-63.
14. Elewski BE. Tinea capitis: A current perspective. J Am Acad Dermatol 2000;42:1-20.
15. Tosti A, Piraccini BM, Lorenzi S. Onychomycosis caused by nondermatophytic molds: Clinical features and response to treatment of 59 cases. J Am Acad Dermatol 2000;42:217-24.
16. Greer DL. Evolving role of nondermatophytes in onychomycosis. Int J Dermatol 1995;34:521-4.
17. Velegraki A, Cafarchia C, Gaitanis G, Iatta R, Boekhout T. Malassezia infections in humans and animals: Pathophysiology, detection, and treatment. PLoS Pathog 2015;11:e1004523.
18. Sobel JD. Recurrent vulvovaginal candidiasis. Am J Obstet Gynecol 2016;214:15-21
19. Sharkey-Mathis PK, Kauffman CA, Graybill JR, Stevens DA, Hostetler JS, Cloud G, et al. Treatment of sporotrichosis with itraconazole. NIAID mycoses study group. Am J Med 1993; 95:279-85.
20. Eicher T, Hauptmann S. “The Chemistry of Heterocycles: Structure, Reactions, Synthesis, and Applications”, 2 nd ed. John Wiley and Sons; 2003.
21. Ghannoum MA, Rice LB. “Antifungal agents: mode of action, mechanisms of resistance, and correlation of these mechanisms with bacterial resistance”. Clin Microbiol Rev 1999; 12:501.
22. Gubbins PO: The systemically acting azoles. In: Wingard J, Anaissie E, eds. Fungal Infections in the Immunocompromised Patient. Boca Raton, Fla: Taylor & Francis Group. 2005; 457-484.
23. Ghannoum MA, Rice LB: Antifungal agents: mode of action, mechanisms of resistance, and correlation of these mechanisms with bacterial resistance. Clin Microbiol Rev 1999; 12:501.
24. Fothergill AW. “Miconazole: A historical perspective”. Expert Rev Anti-infect Ther 2006;4(2):171-5
25. Pfaller MA, Boyken L, Hollis RJ, Messer SA, Tendolkar S, Diekema DJ. “In vitro susceptibilities of clinical isolates of Candida species, Cryptococcus neoformans, and Aspergillus species to itraconazole: global survey of 9,359 isolates tested by clinical and laboratory standards institute broth microdilution methods”. J Clin Microbiol 2005;43:3807.
26. Chapman SW, Daniel CR 3rd. Cutaneous manifestations of fungal infection. Infect Dis Clin North Am 1994;8:879-910.
27. Lilic D, Cant AJ, Abinun M, Calvert JE, Spickett GP. Chronic mucocutaneous candidiasis. I. Altered antigen-stimulated IL-2, IL4, IL-6 and interferon-gamma (IFN-gamma) production. Clin Exp Immunol 1996;105:205-12.
28. Lilic D, Calvert JE, Cant AJ, Abinun M, Spickett GP. Chronic mucocutaneous candidiasis. II. Class and subclass of specific antibody responses in vivo and in vitro. Clin Exp Immunol 1996;105:213-9.
29. David A. Antifungal agents pharmacokinetics and pharmacodynamics of amphotericin, in "Antimicrobial Pharmacodynamics in Theory and Clinical Practice", 2 nd ed. Charles HN, Paul GA, George LD, Takeo M. Eds. CRC Press; 2007. p. 315–26.
30. Johan WM. Pharmacokinetics and Pharmacodynamics of Azoles, in "Antimicrobial Pharmacodynamics in Theory and Clinical Practice", 2 nd ed. Charles HN, Paul GA, George LD, Takeo M. CRC Press; 2007. p. 327–53.
31. Blum RA, D'Andrea DT, Florentino BM. Increased gastric pH and the bioavailability of fluconazole and ketoconazole. Ann Intern Med 1991;114:755.
32. Venkatakrishnan K, Von Moltke LL, Greenblatt DJ. Effects of the antifungal agents on oxidative drug metabolism: clinical relevance. Clin Pharmacokinet 2000;38:111–80.
33. Niwa T, Shiraga T, Takagi A. Effect of antifungal drugs on cytochrome P450 (CYP) 2C9, CYP2C19, and CYP3A4 activities in human liver microsomes. Biol Pharm Bull 2005;28(9):1805-8.
34. Bellmann R. Clinical pharmacokinetics of systemically administered antimycotics. Curr Clin Pharmacol 2007;2:37.
35. Boucher HW, Groll AH, Chiou CC, Walsh TJ. Newer systemic antifungal agents: pharmacokinetics, safety and efficacy. Drugs 2004;64:1997.
36. Zonios DI, Bennett JE. Update on azole antifungals. Semin Respir Crit Care Med 2008;29:198.
37. Miyama T, Takanaga H, Matsuo H. P-glycoprotein-mediated transport of itraconazole across the blood-brain barrier. Antimicrob Agents Chemother 1998;42(7):1738-44.
38. Pappas PG, Kauffman CA, Perfect J. Alopecia associated with fluconazole therapy. Ann Intern Med 1995;123:354.
39. Tan K, Brayshaw N, Tomaszewski K, Troke P, Wood N. Investigation of the potential relationships between plasma voriconazole concentrations and visual adverse events or liver function test abnormalities. J Clin Pharmacol 2006;46(2):235-43.
40. Ahmad SR, Singer SJ, Leissa BG. Congestive heart failure associated with itraconazole. Lancet 2001;357:1766.
41. Gubbins PO, McConnell SA, Amsden JR. Drug interactions associated with antifungal agents. In: Piscitelli SC, Rodvold KA, eds. Drug Interactions in Infectious Diseases. Totowa, NJ: Humana Press; 2005. p. 289-337.
42. Gubbins PO, Amsden JR. Drug-drug interactions of antifungal agents and implications for patient care. Expert Opin Pharmacother 2005;6(13):2231-43.
43. Piscitelli SC, Goss TF, Wilton JH, D’Andrea DT, Goldstein H, Schentag JJ. Effects of ranitidine and sucralfate on ketoconazole bioavailability. Antimicrob Agents Chemother 1991;35(9):1765-71.